

Process intensification in plasma Immunoglobulin G manufacturing using novel technologies providing enhanced productivity

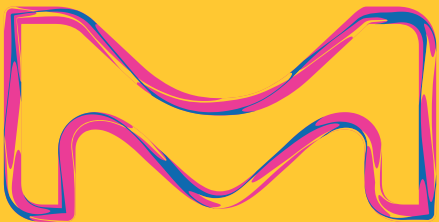
MERCK



Collaboration work with Taipei Medical University, Taipei, Taiwan
(Prof. Thierry Burnouf, College of Biomedical Engineering)

Josephine Cheng
Senior Consultant Strategy Operationalization – traditional modalities – APAC

IPFA/PEI 27th, May 4-6, 2021



The life science business of Merck operates
as MilliporeSigma in the U.S. and Canada.

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Preparation, Separation,
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Agenda

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Introduction – IGG purification

2

Study summary

2

Conclusions

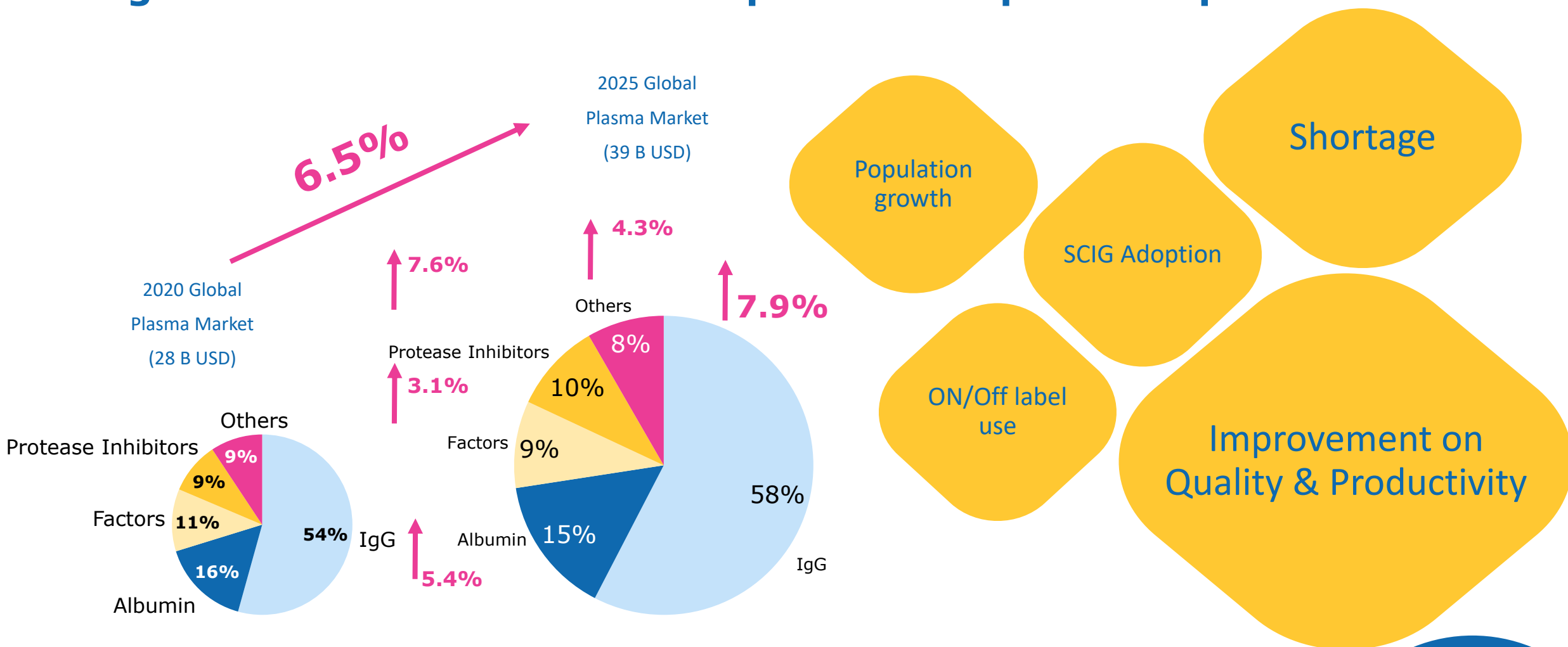
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Immunoglobulin G



High market demands drives exploration in process optimization



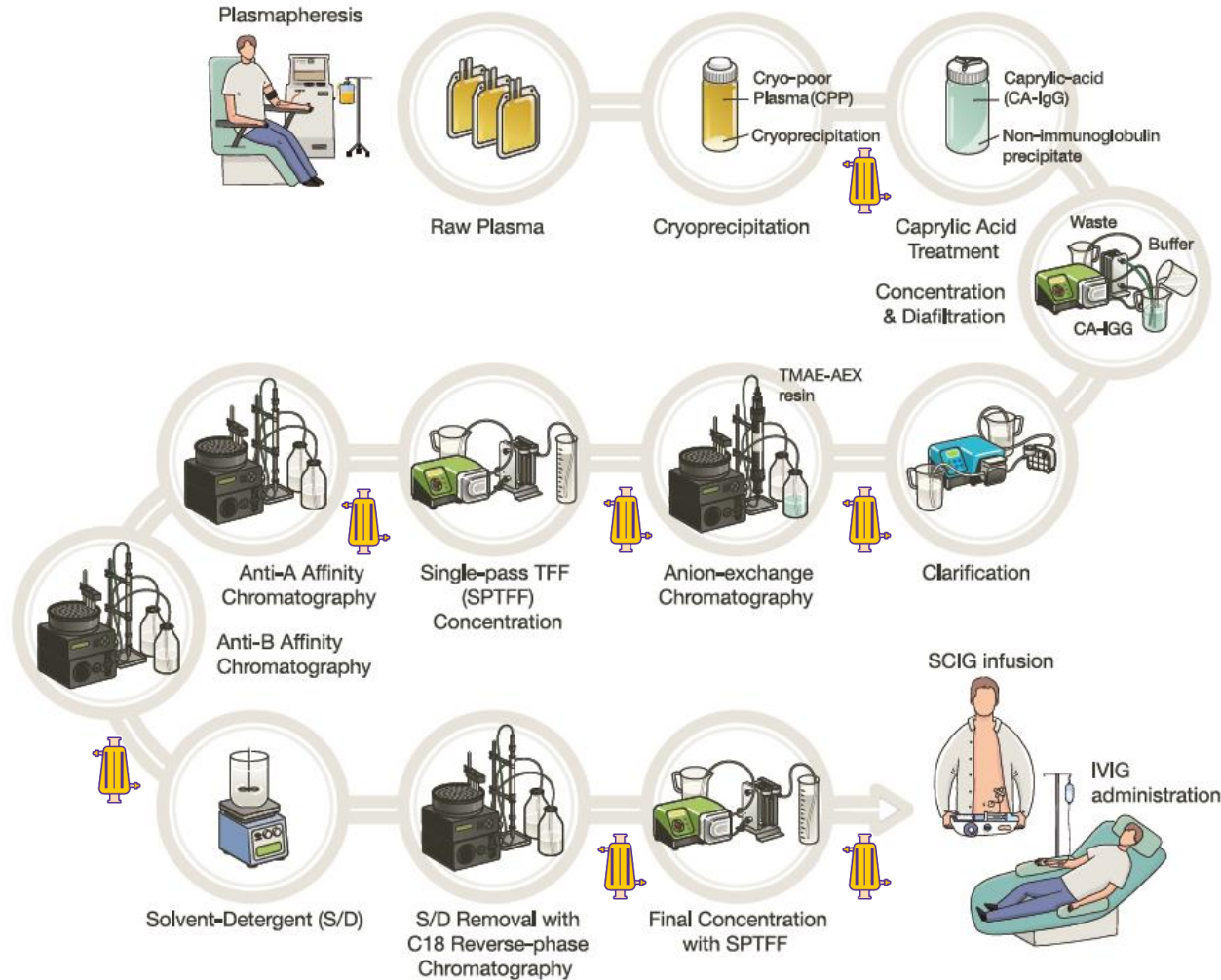
Source: Plasma fractionation markets report 2020, marketsandmarkets.
IPFA 2021 | Josephine Cheng

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From Plasma donation to patient administration

Experimental flow with key steps used in purification

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A generic, easy-to-operate, flowthrough-mode purification process that provides scalable & robust purification with enhanced productivity and quality IGG fitting for therapeutic usage.

Quality criteria:

- Virus safety
- Low IgA & IgM contamination
- Low FXI/XIa
- Lack of Hemolytic effect
- Lack of chemicals used for virus inactivation

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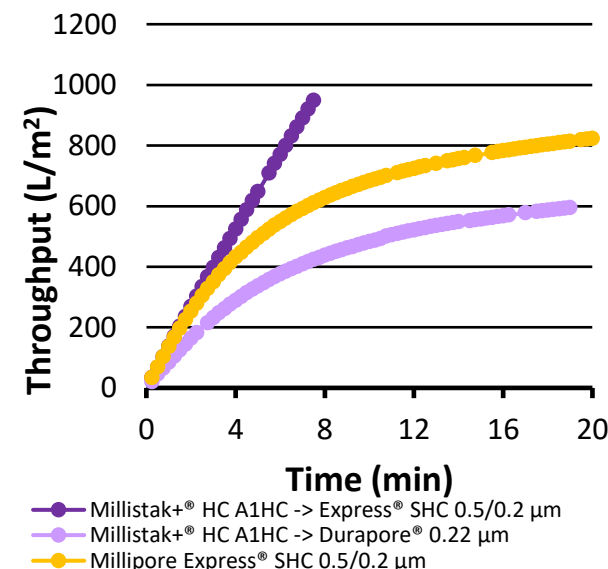
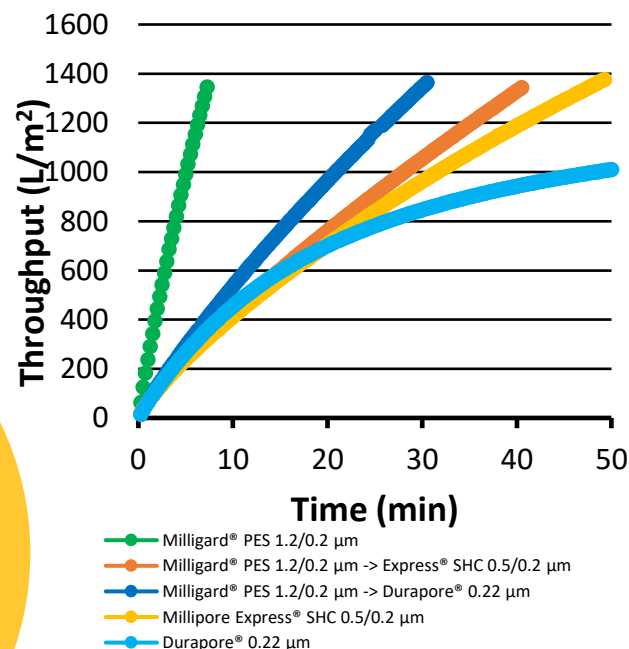
Various intermediate filtration steps reducing bioburden

Prefilters provide significant protection effect to sterile filters and can be used stand alone



Bioburden filters & clarification filters used in the study:

- Millistak+® A1HC
- Milligard® PES 1.2/0.2
- Millipore Express® SHC 0.5/0.2
- Durapore® filter 0.22



Observations:

- w/o prefilter- Millipore Express® SHC generally performs better
- w/ prefilter – sometimes Durapore® performs better
- Prefilter shows significant protecting effect / reducing membrane area needed for sterile filters.
- Millistak® A1HC clarification filter reduced significantly the particles/ precipitation.

Steps	Selected Prefilters (Recovery %)	Selected Sterile Filter	Recovery (%)
Post CA	MPES (100%)	Durapore® filter	> 99
Post UF/DF	Millistak+® A1HC	Millipore Express® SHC	~ 100
Post AEX		Millipore Express® SHC	~ 100
Post 1 st SPTFF	MPES (100%)	Durapore® filter	> 96
Post Anti A/B		Millipore Express® SHC	~ 100
Post C18 resin	MPES (100%)	Millipore Express® SHC	~ 98
Post final SPTFF		Millipore Express® SHC	~ 86

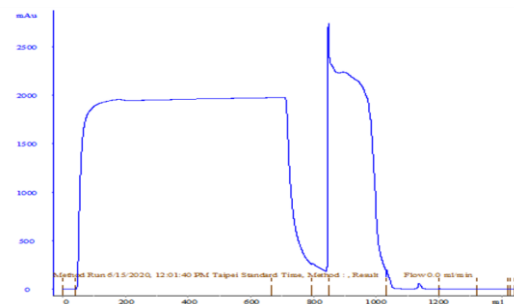
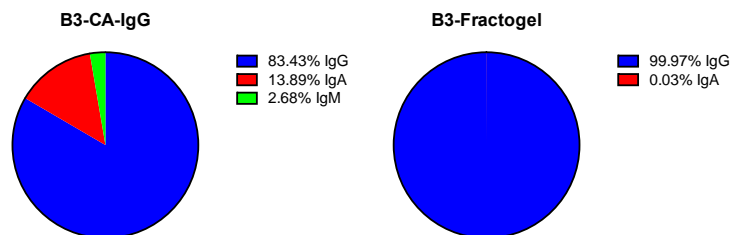
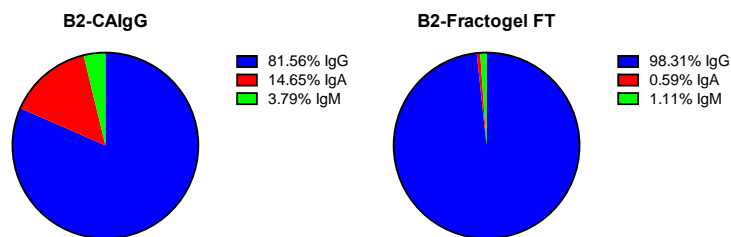
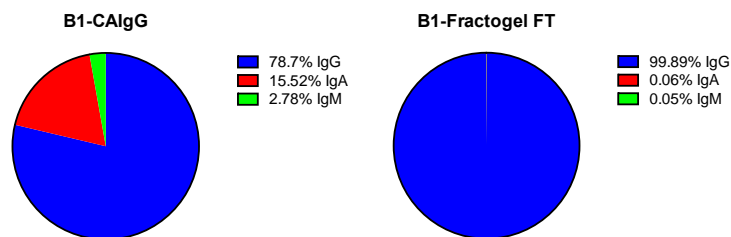
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Anion Exchange chromatography (FT)

Major step to remove IgA & IgM

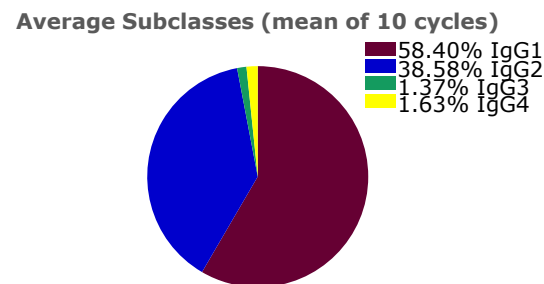
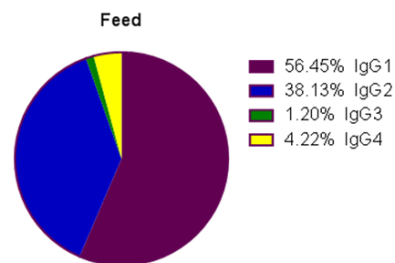
Pilot scale in 3 batches

IgG/IgA/IgM



Small scale in 10 cycles

IgG1/IgG2/IgG3/IgG4



Summary:

1. Optimal pH for loading identified @ pH 6.0
2. Purity IgG avg. 82% to 99% in small and pilot scale.
3. 200 cycles test with standard cleaning conditions confirms the robustness.
4. No changes in IgG subclasses.
5. No thrombogenicity activity detected

Learn More with our webinar:
[Chromatography: Chromatographic strategies for IVIG purification - Part 2](#)

Immunoaffinity chromatography

Robustly reducing the blood-type specific isoagglutinins



Eshmuno® P Anti-A (FT)

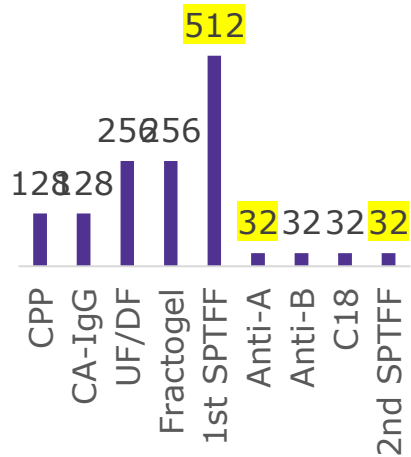


Eshmuno® P Anti-B (FT)

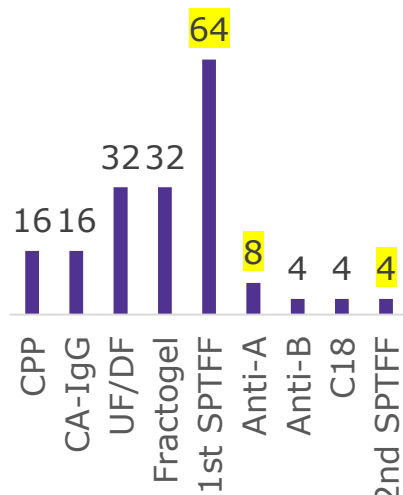


	Group A	Group B	Group AB	Group O
Red blood cell type				
Antibodies in Plasma	 Anti-B	 Anti-A	None	 Anti-A and Anti-B

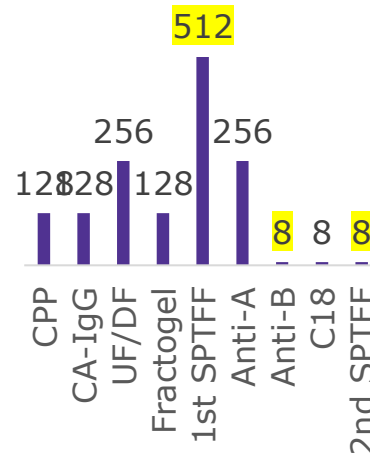
Batch 3 Anti-A titer



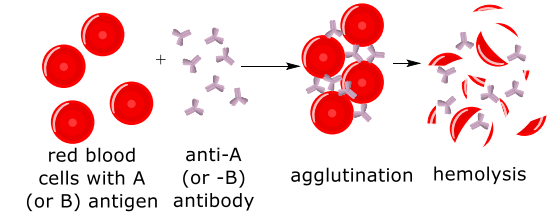
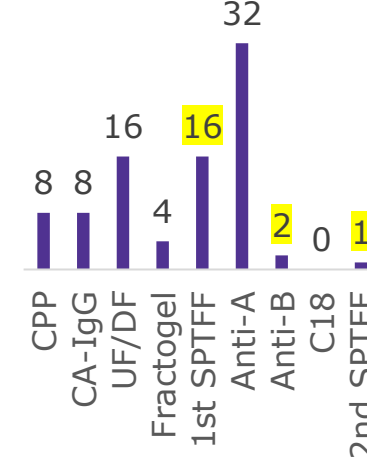
Batch 2 Anti-A titer



Batch 3 Anti-B titer



Batch 2 Anti-B titer



8 to 16 times reduction in Anti-A titer

16 to 32 times reduction in Anti-B titer

*samples tested at 30mg/ml concentration from 1st SPTFF (6X) step to the last step.

Virus Inactivation and removal by C18 RP-chromatography

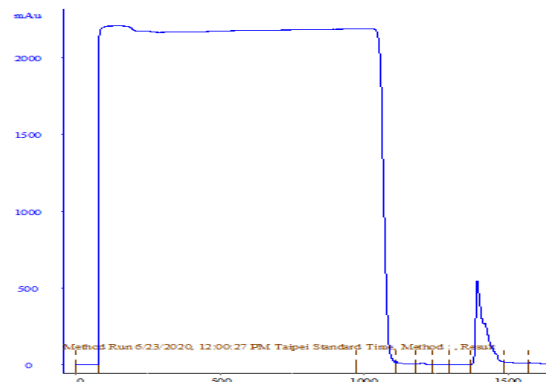


Efficient removal of S/D by Licroprep C18 (40 – 63um) to non-detectable level

A.

Human IgG : 0.3% TnBP + 1% TX100 LRV Results					
Virus	Device	LRV at Incubation Time (min)			
		5	30	60	360
XMuLV	Mobius 1	≥5.5	≥5.3	≥5.3	≥5.4
	Mobius 2	≥5.5	≥5.3	≥5.3	≥5.5
BVDV	Mobius 1	≥4.5	≥4.4	≥4.6	≥4.5
	Mobius 2	≥4.4	≥4.6	≥4.4	≥4.5

B.



C.

Residual TnBP of SD-IgG (Ratio of resin and loaded IgG)	Batch 3 (1mL:6mL)
C18	<1 ppm
SPTFF-5X	<1 ppm

Residual Triton X-100 of SD-IgG (Ratio of resin and loaded IgG)	Batch 3 (1mL:6mL)
C18	<2 ppm
SPTFF-5X	<2 ppm

Summary:

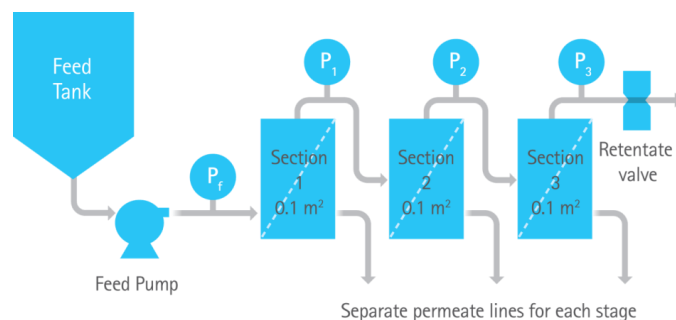
- A. 0.3% TnBP + 1% Triton X-100 provides > 4-5 LRV in time as short as 5 minutes.
- B. Typical chromatography for FT mode S/D-IGG running through C18 column.
- C. TnBP residual tested on GC-MS with results < 1ppm, Triton X-100 residual tested by HPLC with results < 2ppm, showing a robust removal with the loading quantity defined (6ml S/D-IGG/ml C18 packed resin), with capacity tested at 14 ml S/D-IGG/ml C18 resin)

Learn More with our webinar: [Solvent Detergent Viral Inactivation using S.U Technology in Blood Fractionation Processes](#)

*Table A source: Hsieh YT, Mullin L, Greenhalgh P, Cunningham M, Goodrich E, et al.: Single-use technology for solvent/detergent virus inactivation of industrial plasma products. *Transfusion* 2016; 56: 1384-93.

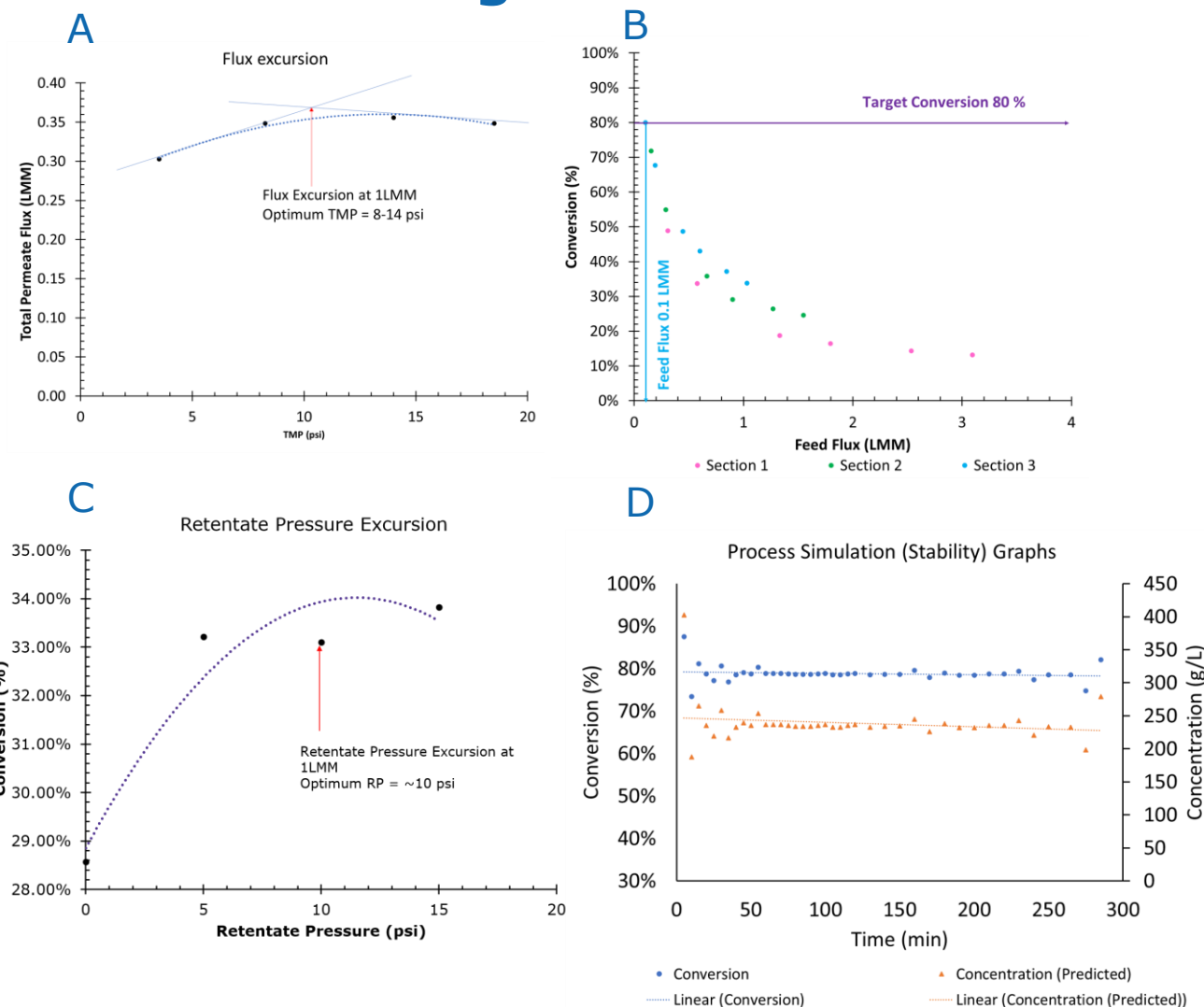
Single-Pass Tangential Flow Filtration

A gentle method for inline concentration & high concentration ultrafiltration



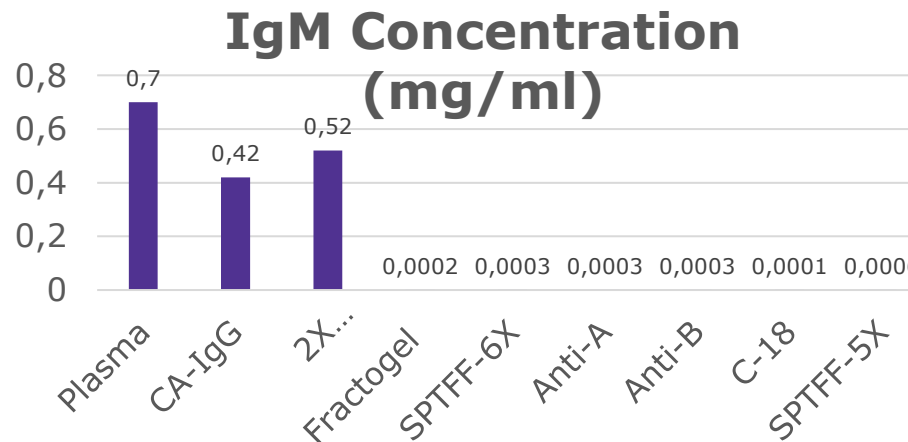
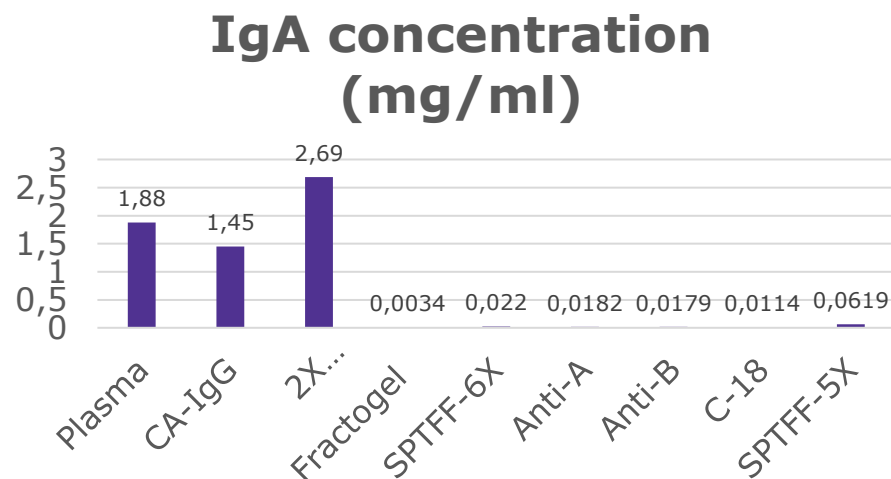
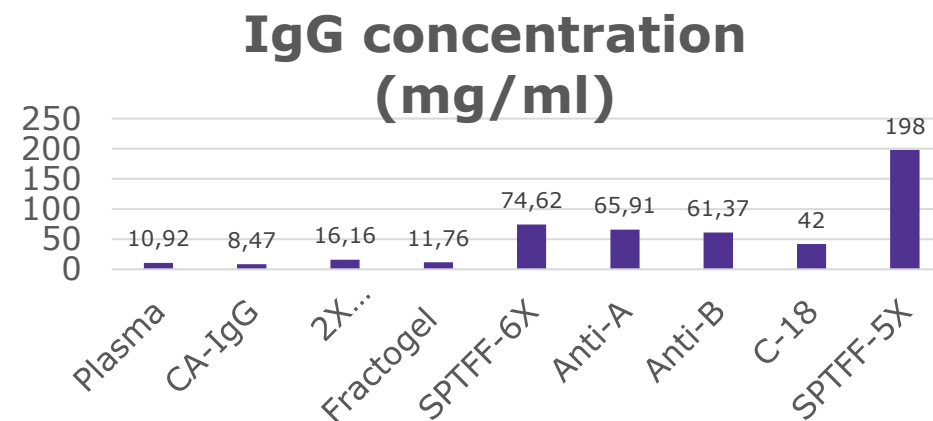
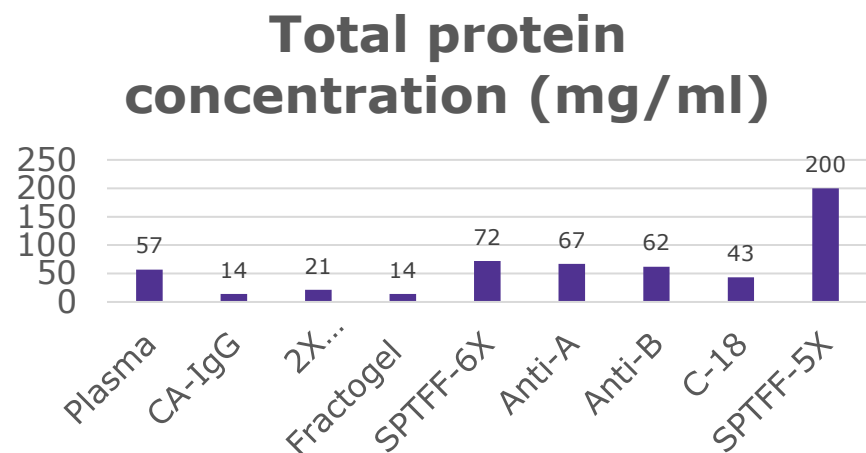
Summary:

1. Sequential optimization to identify parameters to reach target concentration 20%. (200 mg/ml)
2. SPTFF before Eshmuno® P Anti-A/B step reduced loading time, buffer consumed, also improved the AC performance.
3. SPTFF for final concentration successfully provided a gentle (low shear force), scalable, high recovery, and easy-to-operate method for high concentration ultrafiltration.



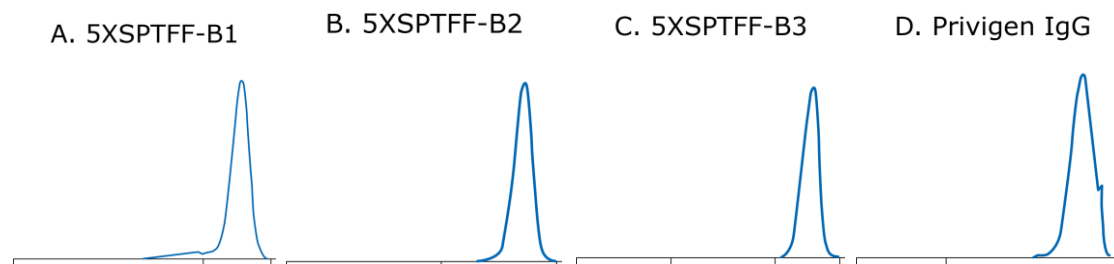
Overall Quality test results

Satisfying concentration effect and IgA/IgG removal ability



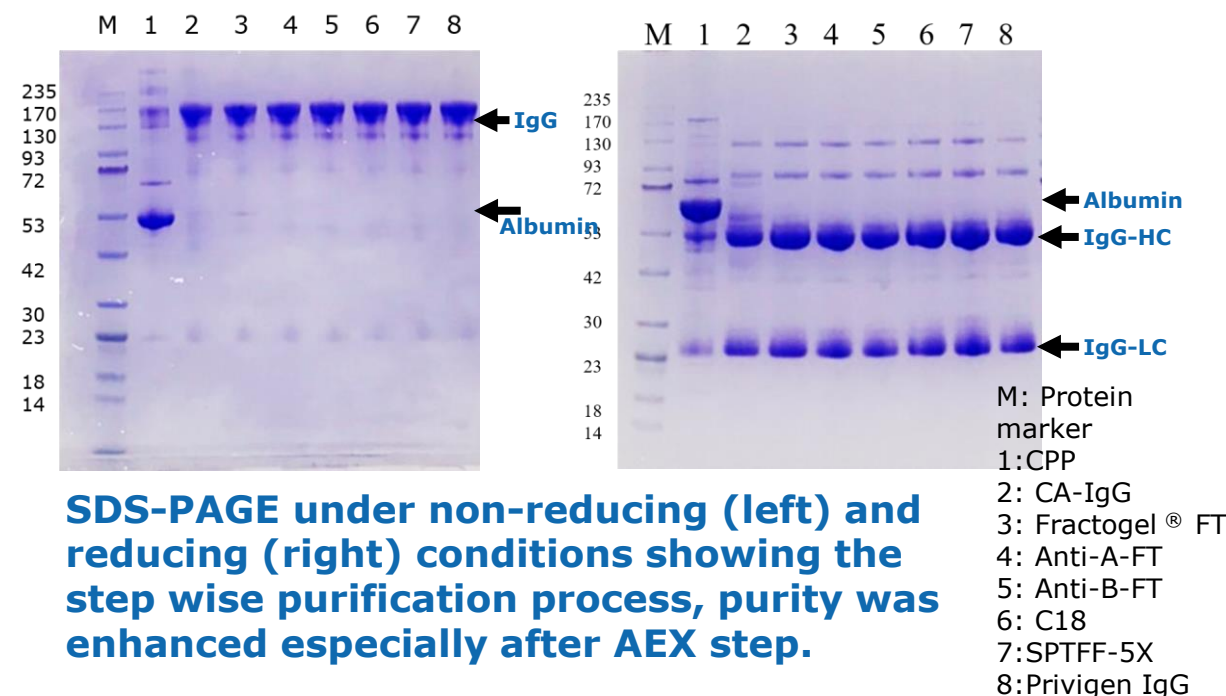
Quality check benchmarking market product

Comparable purity to market product



Fractions	Non-IgG	IgG
Unit	%	%
5XSPTFF-B1	4.8	95.2
5XSPTFF-B2	1.1	98.9
5XSPTFF-B3	0.6	99.4
Privigen® IgG	0.3	99.7

Zone electrophoresis confirmed the high purity of the final IgG. The purity of batch 3 (5X SPTFF) reaches almost 100%.



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*5 µg of protein loaded on to 4-12% Bis-Tris SDS-PAGE

Conclusions



Proof of concept for a generic process with intensified processing:

1. Milligard® PES 1.2/0.2 prefilters effectively reduced the filtration area needed for sterile filters e.g. Millipore Express® SHC or Durapore® filters
2. Clarification using Millistak+® HC A1HC to facilitate downstream purification.
3. Fractogel® TMAE (M) anion exchange chromatography for efficient removal of IgA and IgM, with well maintained IgG subclasses.
4. Eshmuno® P anti-A and Eshmuno® P anti-B chromatography for removal of anti-A and anti-B agglutinins.
5. S/D used in virus inactivation can be efficiently removed using Licroprep® C18 RP chromatography.
6. Use of SPTFF as a mild and robust approach to concentrate IgG to a target of 20% concentration.
7. Recovery from 92 – 100% for each step resulting in an overall process recovery of greater than 70% under a worst-case scenario, with opportunities to improve further with additional optimization.
8. Such flow through methods combined with single-pass TFF technology should be readily scalable, and easy to apply for various IgG products including polyvalent IgG, hyperimmune, or convalescent immunoglobulins.

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Acknowledgements

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Merck:

- Sharon ShangJung Wu
- Karen Waiyu Chan
- Leo Xun Liao
- Xisheng Cao
- Bin Wang

The Millipore logo is located in the bottom right corner, featuring the word "Millipore" in a bold, white, sans-serif font with a registered trademark symbol (®) to its upper right. It is set against a blue, rounded, mountain-like background shape.

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Thank You

Josephine.cheng@merckgroup.com

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